

Note

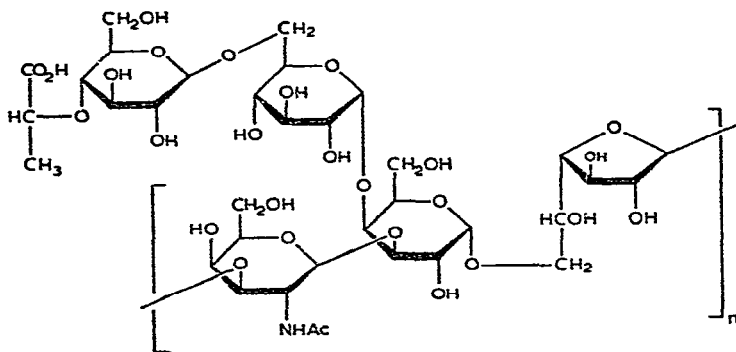
Complete structure of the repeating unit of the O-specific polysaccharide chain of *Shigella dysenteriae* type 3 lipopolysaccharide

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The complete structure of the pentasaccharide repeating-unit of the O-specific polysaccharide chain of the lipopolysaccharide of *Sh. dysenteriae* type 3 has now been established as 1. Previous investigations¹⁻³ revealed all the structural features except the configuration of the glycosidic linkages of the 4-*O*-[(*R*)-1-carboxyethyl]-D-glucose] and D-galactofuranose residues.



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When the disaccharide 6-*O*-{4-*O*-[(*R*)-1-carboxyethyl]-D-glucopyranosyl}-D-glucose, obtained¹ by partial hydrolysis of the polysaccharide with acid, was treated with boron trichloride⁴, a neutral disaccharide was formed together with 4-*O*-[(*R*)-1-carboxyethyl]glucose and glucose, as shown by paper chromatography and electrophoresis. The neutral fraction, obtained after removal (Dowex-1 x8 resin) of the acidic components, was investigated by using a Technicon sugar analyzer and found to contain gentiobiose and glucose. Thus, the 4-*O*-[(*R*)-1-carboxyethyl]-D-glucose residue was β -linked in the polysaccharide. From the optical rotation data in Table I

for the methyl ester of the polysaccharide and the methyl glycosides of the component sugars, it therefore follows that the D-galactofuranose residue also has the β configuration.

TABLE I
OPTICAL ROTATION DATA

Compound	$[\alpha]_D^{25}$ (degrees)	Ref.	$[M]_D^{25}$ (degrees)
Methyl α -D-glucopyranoside	+158.9	6	+308
Methyl α -D-galactopyranoside	+178.8	6	+347
Methyl 2-acetamido-2-deoxy- β -D-galactopyranoside	-12.0	7	-28
Methyl 4-O-[(R)-1-(methoxycarbonyl)ethyl]- β -D-glucopyranoside	+5.0	3	+14
Methyl α -D-galactofuranoside	+104.0	6	+202
Methyl β -D-galactofuranoside	-102.0	8	-198
Methyl ester of polysaccharide, calc. for repeating unit containing an α -D-galactofuranose residue	+90.03		+843
calc. for repeating unit containing a β -D-galactofuranose residue	+47.34		+443
Methyl ester of polysaccharide	+36.5		+342

^aFor aqueous solutions.

Calculation according to Klyne's rule⁵ shows that the molecular rotation (+443°) of the methyl ester of the polysaccharide containing β -D-galactofuranose residues corresponds much more closely to the observed value (+342°) than does the value (+843°) calculated on the basis of α -D-galactofuranose residues.

The above assignment of linkage configuration in the polysaccharide was confirmed by p.m.r. spectroscopy; signals of five anomeric protons at δ 4.40, 4.50, 4.76, 4.94, and 5.06 were observed¹ for a solution of the polysaccharide in D₂O. In accordance with literature data^{9,10}, the anomeric protons of α - and β -hexopyranoside residues resonate at $\delta \sim 5.0$ and ~ 4.50 , respectively. Thus, the signals at δ 4.94 and 5.06 for the polysaccharide correspond to the α -linked residues of D-glucopyranose and D-galactopyranose, whereas the signals at δ 4.40, 4.50, and 4.76 are due to the β -linked residues of 2-acetamido-2-deoxy-D-galactopyranose, 4-O-[(R)-1-carboxyethyl]-D-glucose, and D-galactofuranose.

EXPERIMENTAL

The isolation of oligosaccharides from a partial hydrolysate of the polysaccharide was carried out as previously described¹. Treatment of the acidic disaccharide with boron trichloride was carried out according to the standard procedure⁴. P.c. (descending) was performed on Filtrak FN-11 paper with 1-butanol-pyridine-water (6:4:3). Paper electrophoresis was performed with a 25mM pyridinium acetate buffer

(pH 4.50) at 28 V/cm. Sugars were identified by using a Technicon SC-2 system with a column (25 × 0.6 cm) of DAX4 resin (Durrum, USA), a 0.5M sodium borate buffer (pH 9.0) at 85°, and an elution rate of 60 ml/h. Optical rotations were determined on a Perkin-Elmer polarimeter Model 141.

REFERENCES

- 1 B. A. DMITRIEV, L. V. BACKINOWSKY, V. L. LVOV, N. K. KOCHETKOV, AND I. L. HOFMAN, *Eur. J. Biochem.*, **50** (1975) 539–547.
- 2 N. K. KOCHETKOV, B. A. DMITRIEV, V. L. LVOV, AND L. V. BACKINOWSKY, *Bioorg. Chem. USSR*, **1** (1975) 1238–1240.
- 3 N. K. KOCHETKOV, B. A. DMITRIEV, AND V. L. LVOV, *Carbohydr. Res.*, **54** (1977) 253–259.
- 4 T. G. BONNER AND E. J. BOURNE, *Methods Carbohydr. Chem.*, **2** (1963) 206–207.
- 5 W. KLYNE, *Biochem. J.*, **47** (1950) xli–xlii.
- 6 F. MICHEEL, *Chemie der Zucker und Polysaccharide*, Akademische Verlagsgesellschaft, Leipzig, 1956, pp. 429–431.
- 7 Z. TARASEJEWSKA AND R. W. JEANLOZ, *J. Am. Chem. Soc.*, **80** (1958) 6325–6327.
- 8 J. ANGESTAD AND E. BERNER, *Acta Chem. Scand.*, **8** (1954) 251–254.
- 9 G. M. BEBAULT, Y. M. CHOY, G. G. S. DUTTON, N. FUNNEL, A. M. STEPHEN, AND M. T. YANG, *J. Bacteriol.*, **113** (1973) 1345–1347.
- 10 J. M. VAN VEEN, *J. Org. Chem.*, **28** (1963) 564–566.